

Safety of House Dust Mite Subcutaneous Immunotherapy with a rush and cluster combination protocol in the build-up phase

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Abstract

Background: Conventional and cluster subcutaneous immunotherapy (SCIT) are effective but may be time-consuming. Rush SCIT may offer a more convenient treatment option to patients and be of shorter duration; however, it is also associated with a higher incidence of systemic adverse reactions. Therefore, a combination of protocols between rush and cluster SCIT could have a superior risk-benefit ratio.

Objective: To determine the safety of the combination of rush and cluster HDM-SCIT and to identify the risk factors for local and systemic adverse reactions.

Methods: We retrospectively reviewed the charts of patients who received HDM-SCIT, with rush and cluster combination protocols, at a tertiary care hospital between January 2009 and December 2020. Data were collected at the initial visit (demographic data; underlying allergic disease; current medication; and laboratory investigation results including skin prick test, serum specific IgE (sIgE) levels to aeroallergen, total IgE, and eosinophil count) and follow-up visits (rate and severity of local and systemic adverse reactions).

Results: In total, 698 injections (28 patients) were reviewed. Overall, 13 patients developed systemic adverse reactions, at 3% (21/698) per injection visit. All reactions occurred within 60 minutes. In total, 6 patients experienced large local reactions, at 1.1% (8/698) per injection visit. A high level of sIgE to *D. pteronyssinus* was significantly associated with systemic adverse reactions (HR = 1.02; P = 0.009).

Conclusion: HDM-SCIT with a combination of rush and cluster schedules in the build-up phase could be used as an alternative protocol, given its acceptable systemic adverse reaction rate and shortened duration.

Key words: Allergen, Immunotherapy, House dust mites, Risk factor, Safety

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Introduction

Subcutaneous immunotherapy (SCIT) is a highly effective therapy for allergic diseases, achieving long-term symptom remission.¹ SCIT is indicated for the treatment of severe allergic rhinitis, asthma, and venom anaphylaxis and is currently the only immune-modifying treatment for allergic diseases.^{2,3} House dust mites (HDMs), particularly *Dermatophagoides pteronyssinus* (Dp) and *Dermatophagoides farina* (Df), are regarded as the most important indoor allergens causing allergic sensitization globally, especially in



tropical countries such as Thailand.^{4,5} In addition, they lead to a number of allergic diseases such as allergic rhinitis, atopic asthma, and atopic dermatitis.^{6,7} HDM-SCIT is beneficial to these patients.

The SCIT protocol consists of build-up and maintenance phases. The build-up phase divides the protocol into conventional and accelerated schedules.8 Conventional immunotherapy generally consists of weekly or biweekly allergen injections, which are administered in gradually increasing doses until maintenance doses are reached at around 4 to 6 months, which is quite a long time. Further, this requires frequent hospital visits and possibly time away from school or work, resulting in treatment failure.9 Accelerated build-up schedule involving cluster and rush build-up has been an alternative beneficial treatment method, which enables patients to rapidly achieve the maintenance dose. Cluster build-up usually includes 2-4 sequential injections per day on nonconsecutive days, allowing patients to reach the maintenance dose in 4-8 weeks. Rush build-up involves an increasing dose administered at intervals of 15-60 minutes over the course of 1-3 days until the maintenance dose is reached.¹⁰ However, these methods are also associated with a greater risk of developing systemic responses.

Previous studies on HDM-SCIT have demonstrated that the incidence of systemic adverse reactions (SAEs) ranged from 0.31% to 4.6% per injection (3–20% of patients) for a conventional schedule, whereas this incidence is approximately 0.15-3.5% (7% of patients) for a cluster schedule and is higher at 4.2% per injections (27–35% of patients) for a rush schedule.¹⁰⁻¹⁴

A significant degree of skin test reactivity and a forced expiratory volume in one second (FEV₁) less than 80% of predicted were reported to be significant predictors of systemic reactions with rush immunotherapy to HDMs.⁹ Higher body mass index (BMI) and serum specific IgE (sIgE) levels to HDMs are also reported as risk factors for developing local reactions (LRs).¹⁵

Therefore, combining rush and cluster protocols is preferable with respect to the risk-benefit ratio. However, the safety of this method against HDMs has not been thoroughly investigated. Hence, this study aimed to determine the safety of the combination of rush and cluster HDM-SCIT in the build-up phase and to identify risk factors for local and systemic adverse reactions.

Methods

Study design and study population

This was a retrospective chart review of patients who received HDM-SCIT with a rush and cluster combination protocol in the build-up phase at a tertiary care hospital; from January 2009 to December 2020. The eligibility criteria for HDM-SCIT were as follows: age > 5 years, treatment-resistant allergic disease, defined as patients with moderate-to-severe allergic rhinitis, moderate-to-severe allergic conjunctivitis, moderate-to-severe atopic dermatitis, and/or persistent asthma, that are not well tolerated with optimal avoidance measures and pharmacotherapy, sensitization to HDMs (sIgE level > 0.35 kUA/L), and FEV₁ \geq 70%. Patients who did not complete the study protocol were excluded.

Data was collected from the medical records. Data on demographics (sex, age, underlying disease, onset of diagnosis, duration of treatment prior to receiving SCIT, and atopy comorbidity); total serum IgE level; serum eosinophil count; current medication; spirometry results; and aeroallergen serum sIgE levels to *D. pteronyssinus*, *D. farinae*, cockroach, bermuda, cat pelt, and dog pelt were collected at the initial visit. Follow-up data were also recorded, focusing on SAEs (onset and severity of the reaction) and large local reactions (LLRs). sIgE levels for aeroallergens were assessed using ImmunoCap. Sensitization to aeroallergens was defined as a sIgE level > 0.35 kUA/L.

Allergen extract and rush and cluster combination protocol

HDM-SCIT was performed using an HDM preparation (ALK-Abello, Port Washington, NY, USA) containing a 50:50 mixture of *D. pteronyssinus* and *D. farinae* extracts. The HDM extract used for maintenance immunotherapy was 500 AU (0.5 mL/dose), which was a mixture of the 250 AU/each allergen extracts.

As part of the HDM-SCIT rush protocol, all patients were admitted to the hospital for 3 days (three injections per day for three consecutive days) and were administered prednisolone (1 mg/kg/day, max 40 mg) and an antihistamine (cetirizine 5 mg or loratadine 5 mg) 1 hour before the first injection. Subsequently, they visited the allergy clinic once weekly for the HDM-SCIT cluster protocol, in which the patient received two injections each day. Accordingly, the rush and cluster combination protocol consisted of 7 weeks and 21 injections (**Table 1**). A physician, nurse practitioner, or physician assistant was present during immunotherapy administration and during the observation period. Patients were observed for 30 minutes between the doses and after the final dose of the day.

			Concentration of allergen extract	Volume (mL)
	Week 1	Day 1	1:1000	0.1
			1:1000	0.2
			1:1000	0.4
dule		Day 2	1:100	0.1
Rush schedule			1:100	0.3
Rush			1:100	0.5
		Day 3	1:10	0.1
			1:10	0.2
			1:10	0.3

Table 1. Rush and cluster combination protocol.

Table 1. (Continued)

		Concentration of allergen extract	Volume (mL)
	Week 2	1:10	0.35
	week 2	1:10	0.4
	Week 3	1:10	0.5
	week 3	1:1	0.1
ule	Week 4	1:1	0.15
sched	week 4	1:1	0.2
Cluster schedule	Week 5	1:1	0.25
Clu	Week 5	1:1	0.3
		1:1	0.35
	Week 6	1:1	0.4
	Week 7	1:1	0.45
	week /	1:1	0.5

Evaluation of reactions

All systemic reactions were assessed based on the World Allergy Organization Subcutaneous Immunology Systemic Reaction Grading System. Large local reactions (LLRs) were assessed according to the size of the local swelling (wheal), and those with a mean diameter of 25 mm were considered LLRs.³

Sample size calculations

We calculated the sample size for the detected rate of systemic adverse reaction of HDM-SCIT in the built-up phase, using a formula for estimating the infinite population proportion (P; rate of adverse reaction in rush protocol = 0.04).¹⁰ The overall sample size was calculated as 60 injections.

Statistical analysis

Demographic data are presented as the mean \pm standard deviation or median for continuous data and as number (%) for categorical data. The number and severity of systemic and local adverse reactions, systemic reaction rate per injection visit, and local reaction rate per the number of injections were determined. Cox regression analysis was used for univariate and multivariate analyses. Variables, including baseline characteristics and laboratory findings, were used to estimate the risk factors associated with systemic reactions. All statistical analyses were performed using a standard software package (Stata version 13.0; StataCorp). P < 0.05 was considered significant.

Ethics statement

This study was approved by the Ethics Committee and Institutional Review Board of the Faculty of Medicine, Prince of Songkla University (IRB number is 64-118-01-1). Safety of House Dust Mite immunotherapy

Results

Patient characteristics

A total of 28 patients (698 injections) who underwent rush and cluster combination subcutaneous immunotherapy for HDM were evaluated (**Table 2**). The mean age was 14.7 years, and 16 children (57.1%) were male. The most common diseases indicated for HDM-SCIT were atopic dermatitis (35.7%), asthma (32.1%) and allergic rhinitis (25%), and allergic rhinitis (81.8%) was the most common comorbidity. The median sIgE to *D. pteronyssinus* was 73 kU/L, and the mean sIgE to *D. farinae*, 48.6 kU/L. The mean FEV₁ was 92%, and 46% of patients received more than 400 mcg of inhaled corticosteroid once daily. The mean serum total IgE level was 2,010.2 kU/L, and the mean eosinophil count was 563.3 mm³.

Table 2. Baseline characteristics.

Characteristic	Total (N = 28)	
Age (years), median (IQR)	14.7 (12.6,16)	
Sex (male), n (%)	16 (57.1)	
Diseases that are indicated for immunotherapy, n (%)		
Asthma	9 (32.1)	
Allergic rhinitis	7 (25.0)	
Atopic dermatitis	10 (35.7)	
Severe allergic conjunctivitis	1 (3.6)	
Chronic spontaneous urticaria	1 (3.6)	
Comorbid of allergic diseases, n (%)	22 (78.6)	
Asthma	5 (22.7)	
Allergic rhinitis	18 (81.8)	
Atopic dermatitis	3 (13.6)	
Allergic conjunctivitis	1 (4.5)	
Onset of diagnosis (month), median (IQR)	97.4 (67.6-132.1)	
Duration of treatment (month), median (IQR)	34.5 (14.9-54.7)	
Specific serum IgE, kU/L (%)		
Dermatophagoides. pteronyssinus, median (IQR)	73 (19.5, 100)	
Dermatophagoides farinae, mean (SD)	48.6 (42)	
Cockroach, median (IQR)	3.3 (13.1)	
Cat pelt, mean (SD)	11.4 (29.2)	
Bermuda, mean (SD)	2.6 (10)	
Dog pelt, mean (SD)	0.3 (0.3)	
Pulmonary function test, mean (SD)		
FVC	95.7 (10.7)	
FEV ₁	92 (16.5)	
FEV ₁ /FVC	96.7 (9.9)	
FER _{25-75%}	83 (19.3)	
PEFR	84.7 (19.6)	



Table 2. Baseline characteristics.

Characteristic	Total (N = 28)	
Total IgE (kU/L), mean (SD)	2,010.2 (2,567.7)	
Eosinophil number (mm ³), mean (SD)	536.3 (573)	
Omalizumab, n (%)	4 (14)	
Daily dose of inhaled corticosteroid, n (%)	13 (46.4)	
0-200	1 (7.7)	
200-400	6 (46.2)	
> 400	6 (46.2)	
Daily dose of intranasal corticosteroid, n (%)	25 (89.3)	
0-50	5 (20)	
50-100	14 (56)	
> 100	6 (24)	

Abbreviation: IQR, interquartile range; SD, standard deviation; FEV₁, forced expiration in 1 second; FVC, Forced Vital Capacity; FEF_{25-75%}, forced expiratory flow at 25 – 75% of FVC, PEFR, Peak expiratory flow rate

Incidence and severity of adverse reactions

In total, 13 patients developed systemic adverse reactions, 3% (21/698) per injection. Among them, 6 patients developed LLRs, 1.1% (8/698) per injection. The most common adverse reactions were grade II systemic reactions (1.4%). The incidence of systemic adverse reactions was higher in the rush schedule (2.3%); grade I systemic reactions occurred in 1.1%. LLRs were also more common in the rush schedule than in the cluster schedule (0.9% vs 0.3%) (**Table 3**). Almost all systemic reactions occurred within the first 60 minutes of injection. The characteristics, including prescribed medication of the patients who developed systemic adverse reactions, are shown in **Table 4**.

Risk factors associated with SAEs and LLRs

Univariate analysis showed that serum sIgE to Dp was significantly associated with the development of SAEs (HR = 1.01; P = 0.02). Meanwhile, age and serum sIgE to cockroaches were significantly associated with decreased SAEs with HR = 0.93; P = 0.03 and HR = 0.39; P = 0.04, respectively. FEV₁ and receiving omalizumab decreased the incidence of SAEs; however, this difference was insignificant. In terms of LLRs, univariate analysis revealed that serum sIgE to Df was significantly associated with LLRs (HR = 1.02; P = 0.04) (Table 5).

Number of reactions classified Number of overall by schedule of build-up phase **Reaction grade** reactions (N = 698)**Rush schedule Cluster schedule** Systemic reaction, n (%) 21 (3.0) 16 (2.3) 5 (0.7) Grade 1, n (%) 8 (1.1) 8 (1.1) 0(0)Grade 2, n (%) 10 (1.4) 5 (0.7) 5 (0.7) Grade 3, n (%) 3 (0.4) 3(0.4)0(0)LLR, n (%) 8 (1.1) 6 (0.9) 2 (0.3)

Table 3. Reactions from combination rush and cluster HDM-SCIT protocol.

Abbreviation: LLR, large local reaction

Table 4. Type, severity, and timing of systemic reaction during combination rush and cluster HDM-SCIT protocol.

No		Dose of HDI	M-SCIT		Detail of	systemic adverse reaction	
	Schedule	Concentration of allergen extract	Volume (mL)	Onset (minutes)	Severity grade of systemic reaction	Clinical presentation	Treatment
1	Rush	1:100	0.5	30	3	Generalized urticaria, expiratory wheezing	Adrenaline, CPM, hydrocorticosteroid
2	Rush	1:10	0.2	60	3	Generalized urticaria, angioedema, wheezing	Adrenaline, CPM, hydrocorticosteroid
3	Rush	1:100	0.3	60	1	Generalized urticaria	СРМ
4	Rush	1:10	0.3	30	2	Generalized urticaria	СРМ
5	Rush	1:10	0.2	25	2	Generalized urticaria, expiratory wheezing	Adrenaline, CPM, hydrocorticosteroid, Ventolin



No		Dose of HDN	M-SCIT		Detail of	f systemic adverse reaction		
	Schedule	Concentration of allergen extract	Volume (mL)	Onset (minutes)	Severity grade of systemic reaction	Clinical presentation	Treatment	
6	Cluster	1:1	0.1	40	2	Expiratory wheezing	Ventolin, Prednisolone	
7	Rush	1:100	0.5	30	1	Generalized urticaria	СРМ	
8	Rush	1:10	0.5	20	1	Generalized urticaria	Cetirizine	
9	Rush	1:100	0.5	40	2	Expiratory wheezing, itching	CPM, Ventolin	
10	Rush	1:1000	0.1	30	1	Rhinorrhea, nasal congestion	Cetirizine	
11	Rush	1:100	0.5	30	1	Generalized urticaria	СРМ	
12	Rush	1:10	0.2	30	2	Generalized urticaria, expiratory wheezing	Adrenaline, CPM, hydrocorticosteroid, Ventolin	
13	Rush	1:10	0.25	30	1	Generalized urticaria	СРМ	
14	Cluster	1:1	0.3	30	2	Cough, nasal congestion, expiratory wheezing	Ventolin	
15	Rush	1:100	0.5	90	1	Generalized urticaria, chest tightness, cough	СРМ	
16	Cluster	1:1	0.2	180	2	Local urticaria, chest tightness	Cetirizine, Ventolin	
17	Cluster	1:1	0.2	120	2	Expiratory wheezing, nasal congestion	Ventolin, Prednisolone	
18	Cluster	1:1	0.5	240	2	Local urticaria, chest tightness	Cetirizine, Ventolin	
19	Rush	1:10	0.3	60	3	Cough, nasal congestion, expiratory wheezing	Adrenaline, CPM, hydrocorticosteroid, Ventolin	
20	Rush	1:10	0.2	60	1	Generalized urticaria, nasal congestion	Adrenaline, CPM, hydrocorticosteroid	
21	Rush	1:10	0.2	60	2	Generalized urticaria, nasal congestion, chest tightness	Adrenaline, CPM, hydrocorticosteroid	

Table 4. (Continued)

Table 5. Univariate analysis for risk factors associated with systemic and large local reactions.

Tastan		SAEs		LLRs			
Factors	HR	95%CI	p Value	HR	95%CI	<i>p</i> Value	
Age (years)	0.93	0.87-0.99	0.03	1.05	0.98-1.12	0.19	
Sex (male)	1.27	0.40-4.04	0.68	0.45	0.09-2.19	0.32	
Asthma	1.13	0.28-4.54	0.87	3.11	0.30-32.20	0.34	
Degree of skin test to Dp	1.05	0.84-0.30	0.64	1.23	0.37-4.16	0.74	
Degree of skin test to Df	1.15	0.88-1.52	0.30	1.12	0.57-2.19	0.75	
sIgE to Dp	1.01	1.00-1.03	0.02	1.01	0.99-1.03	0.18	
sIgE to Df	1.00	0.99-1.02	0.76	1.02	1.00-1.04	0.04	
sIgE to Cockroach	0.39	0.16-0.94	0.04	0.98	0.95-1.02	0.37	
FEV ₁	0.96	0.92-1.01	0.10	0.94	0.86-1.02	0.14	
Omalizumab	0.49	0.08-3.10	0.45	3.00	0.85-15.64	0.19	

Abbreviation: HR, hazard ratio; SAEs, systemic adverse reactions; LLRs, large local reactions; Dp, *Dermatophagoides pteronyssinus*; Df, *Dermatophagoides farina*; sIgE, Specific IgE; FEV₁, forced expiration in 1 second.



To show		SAEs		LLRs			
Factors	HR	95%CI	<i>p</i> Value	HR	95%CI	<i>p</i> Value	
Age (years)	0.96	0.89 - 1.02	0.19	1.04	0.99 - 1.09	0.08	
Sex (male)	1.43	0.61 - 3.37	0.42	1.62	0.37 - 7.04	0.52	
sIgE to Dp	1.02	1.01 - 1.03	0.009				
sIgE to Df				1.02	1.00 - 1.03	0.02	
sIgE to Cockroach	0.29	0.58 - 1.53	0.15				

Table 6. Multivariate analysis for risk factors associated with systemic and large local reactions.

Abbreviation: HR, hazard ratio; SAEs, systemic adverse reactions; LRs, large local reactions; sIgE, Specific IgE.; Dp, Dermatophagoides pteronyssinus; Df, Dermatophagoides farina;

Multivariable analysis revealed that only serum sIgE to Dp was significantly associated with the development of SAEs (HR = 1.02; 95%CI: 1.01-1.03, P = 0.009), while serum sIgE to Df (kU/L) was only significantly associated with the development of LLR. (HR = 1.02; 95%CI: 1.00-1.03, P = 0.25) (**Table 6**).

Other factors such as age, asthma, allergic rhinitis, degree of skin test reactivity, mean serum total IgE level, the mean eosinophil count, and dose of inhaled corticosteroid in asthmatic patients were not associated with the development of SAEs or LLRs.

Discussion

The safety of a combination of cluster and rush protocols in the build-up phase for HDM-SCIT is yet to be clarified. In this study, the rate of systemic reactions was 3% per injection visit. Most of the adverse systemic reactions were mild to moderate and occurred within 60 minutes. Furthermore, the rate of LLR was 1.1% per injection visit.

A recent systematic review showed that systemic reactions occur in 0.05-3.2% and 0.15-3.3% per injection for conventional and cluster aeroallergen immunotherapy, respectively.⁹ In terms of systemic reactions to HDM-SCIT, previous studies have reported that its incidence ranged from 0.31% to 4.6% and from 0.15% to 3.5% per injection for conventional and cluster schedules, respectively. However, this incidence was greater for rush schedules, reaching 4.2% per injection. This study's systemic reactions rate is consistent with those reported for the conventional and cluster technique.¹⁰⁻¹⁴ Moreover, the combination protocol has the advantage of a reduced number of clinic visits, saving the patient time and reducing costs, ultimately increasing the convenience of this treatment.

The rates of systemic reactions in rush immunotherapy (RIT) range from 27% to 100% for patients treated with RIT without premedication and from 7.2% to 27% for patients treated with premedication.¹⁶⁻¹⁸ This indicates that premedication decreases the reaction rate. Many previous studies showed that pretreatment with second-generation antihistamines could reduce the rates of SAEs for accelerated build-up schedules.¹⁹⁻²¹ Moreover, some studies found that omalizumab significantly reduced the rate of SAEs to rush immunotherapy.^{22,23} In this study, all patients were administered antihistamine 1 hour before injection,

and 14% received omalizumab. However, we did not find any significant association between omalizumab and SAEs, possibly because of the small number of patients with asthma diagnoses and the exclusions of people who had FEV_1 less than 70%. In this study, higher systemic reactions occurred during the rush period, indicating that care should be taken during this period. However, most systemic reactions were only of mild to moderate severity (grades 1-3). The patient's symptoms improved with medication, as shown in **Table 4**, and there was no need for intensive unit care; no patient died. Our findings support that patients should be hospitalized during the rush period for close monitoring and premedication before HDM-SCIT injection to reduce the risk of systemic adverse reactions.

One of the main reasons for poor SCIT adherence is the inconvenience of coming to the clinic to receive allergy injections, especially in the build-up phase.^{24,25} Therefore, it has been suggested that shortening the treatment schedules by reducing the number of injections could indirectly improve the adherence rate.²⁶ Compared to other cluster protocols, our protocol helps decrease the cluster period time as we had previously implemented a rush schedule. Accordingly, combining cluster and rush schedules in the build-up phase of the HDM-SCIT protocol can balance the risks and benefits of treatment.

Our results showed that a high level of sIgE to Dp was significantly associated with SAEs, and a high level of sIgE to Df was significantly associated with LLRs. This finding suggests that a high level of HDM sensitization may be associated with systemic reactions to SCIT, in accordance with the findings of other studies that have demonstrated the degree of skin test reactivity as a predictor of systemic reactions during immunotherapy. DaVeiga et al showed that 3-4+ positive skin tests are associated with systemic reactions in aeroallergen immunotherapy (OR: 5.8, 95%CI: 1.2-27.6).²⁷ Parmiani et al also found an association between the degree of skin test reactions in inhalant SCIT.²⁸

There are a number of limitations mentioned in this study. First, since this study is a single-center observational retrospective study and had a small patient population, additional clinical studies involving HDM-SCIT patients with a larger patient population might be required to confirm the safety of the rush and cluster combination protocol. Second, as prednisolone and an antihistamine were administered to



all patients as premedication, this study is unable to assess whether premedication has any effect on reducing SAEs. In addition, regarding the exclusion of patients with FEV_1 less than 70% in this study, it may have an impact on the assessment of the association between omalizumab and SAEs.

Conclusion

HDM-SCIT with a combination of rush and cluster schedules in the build-up phase showed an acceptable systemic adverse reaction rate and shortened duration, indicating that it could be used as an alternative protocol.

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Conflicts of interest

The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article

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This was an unfunded study.

Authors' contributions

- AY, VK designed the study, performed the analysis and manuscript preparation.
- NJ designed the study and performed the data collection.
- PS designed the study and performed the analysis.
- All authors have read and approved the final manuscript.

References

- Jutel M, Kosowska A, Smolinska S. Allergen immunotherapy: Past, present, and future. Allergy Asthma Immunol Res. 2016;8:191-7.
- Alvaro-Lozano M, Akdis CA, Akdis M, Alviani C, Angier E, Arasi S, et al. EAACI allergen immunotherapy user's guide. Pediatr Allergy Immunol. 2020;31:1-101.
- Cox L, Nelson H, Lockey R, Calabria C, Chacko T, Finegold I, et al. Allergen immunotherapy: a practice parameter third update. J Allergy Clin Immunol. 2011;127:S1-55.
- Assarehzadegan M-A, Shakurnia A, Amini A. The most common aeroallergens in a tropical region in Southwestern Iran. World Allergy Organ J. 2013;6:7.
- Oncham S, Udomsubpayakul U, Laisuan W. Skin prick test reactivity to aeroallergens in adult allergy clinic in Thailand: a 12-year retrospective study. Asia Pac Allergy. 2018;8.
- Sporik R, Holgate ST, Platts-Mills TA, Cogswell JJ. Exposure to house-dust mite allergen (Der p I) and the development of asthma in childhood. A prospective study: A prospective study. N Engl J Med. 1990;323:502-7.
- Calderón MA, Linneberg A, Kleine-Tebbe J, De Blay F, Hernandez Fernandez de Rojas D, Virchow JC, et al. Respiratory allergy caused by house dust mites: What do we really know? J Allergy Clin Immunol. 2015;136:38-48.

- Cox L, Li JT, Nelson H, Lockey R. Allergen immunotherapy: A practice parameter second update. J Allergy Clin Immunol. 2007;120:S25-85.
- Cox L. Accelerated immunotherapy schedules: review of efficacy and safety. Ann Allergy Asthma Immunol. 2006;97:126-37; quiz 137-40, 202.
- Calabria CW. Accelerated immunotherapy schedules. Curr Allergy Asthma Rep. 2013;13:389-98.
- Schubert R, Eickmeier O, Garn H, Baer PC, Mueller T, Schulze J, et al. Safety and immunogenicity of a cluster specific immunotherapy in children with bronchial asthma and mite allergy. Int Arch Allergy Immunol. 2009;148:251-60.
- 12. Uchida T, Nakagome K, Iemura H, Naito E, Miyauchi S, Uchida Y, et al. Clinical evaluation of rush immunotherapy using house dust mite allergen in Japanese asthmatics. Asia Pac Allergy. 2021;11:e32.
- Kim ME, Kim JE, Sung JM, Lee JW, Choi GS, Nahm DH. Safety of accelerated schedules of subcutaneous allergen immunotherapy with house dust mite extract in patients with atopic dermatitis. J Korean Med Sci. 2011;26:1159-64.
- 14. Calabria CW, Cox L. Accelerated immunotherapy schedules and premedication. Immunol Allergy Clin North Am. 2011;31:251-63, ix.
- Yang Y, Ma D, Huang N, Li W, Jiang Q, Wang Y, et al. Safety of house dust mite subcutaneous immunotherapy in preschool children with respiratory allergic diseases. Ital J Pediatr. 2021;47:101.
- Lilja G, Sundin B, Graff-Lonnevig V, Hedlin G, Heilborn H, Norrlind K, et al. Immunotherapy with cat- and dog-dander extracts. IV. Effects of 2 years of treatment. J Allergy Clin Immunol. 1989;83:37-44.
- Hejjaoui A, Dhivert H, Michel FB, Bousquet J. Immunotherapy with a standardized Dermatophagoides pteronyssinus extract. IV. Systemic reactions according to the immunotherapy schedule. J Allergy Clin Immunol. 1990;85:473-9.
- Portnoy J, Bagstad K, Kanarek H, Pacheco F, Hall B, Barnes C. Premedication reduces the incidence of systemic reactions during inhalant rush immunotherapy with mixtures of allergenic extracts. Ann Allergy. 1994;73:409-18.
- 19. Ohashi Y, Nakai Y, Murata K. Effect of pretreatment with fexofenadine on the safety of immunotherapy in patients with allergic rhinitis. Ann Allergy Asthma Immunol. 2006;96:600-5.
- Nielsen L, Johnsen CR, Mosbech H, Poulsen LK, Malling HJ. Antihistamine premedication in specific cluster immunotherapy: a double-blind, placebo-controlled study. J Allergy Clin Immunol. 1996; 97:1207-13.
- 21. Berchtold E, Maibach R, Müller U. Reduction of side effects from rush-immunotherapy with honey bee venom by pretreatment with terfenadine. Clin Exp Allergy. 1992;22:59-65.
- 22. Casale TB, Busse WW, Kline JN, Ballas ZK, Moss MH, Townley RG, et al. Omalizumab pretreatment decreases acute reactions after rush immunotherapy for ragweed-induced seasonal allergic rhinitis. J Allergy Clin Immunol. 2006;117:134-40.
- Casale TB, Kline JN, Busse WW, Ballas ZK, Mokhtarani M, Seyfert-Margolis V, et al. Omalizumab pretreatment prevents allergic reactions due to rush immunotherapy (RIT). J Allergy Clin Immunol. 2005;115:S65.
- 24. Reisacher WR, Visaya JM. Patient adherence to allergy immunotherapy. Curr Opin Otolaryngol Head Neck Surg. 2013;21:256-62.
- Cox LS, Hankin C, Lockey R. Allergy immunotherapy adherence and delivery route: location does not matter. J Allergy Clin Immunol Pract. 2014;2:156-60.
- Pfaar O, van Twuijver E, Hecker H, Boot JD, van Ree R, Klimek L. Accelerated up-dosing of subcutaneous immunotherapy with a registered allergoid grass pollen preparation. Int Arch Allergy Immunol. 2013;160:420-4.
- DaVeiga SP, Liu X, Caruso K, Golubski S, Xu M, Lang DM. Systemic reactions associated with subcutaneous allergen immunotherapy: timing and risk assessment. Ann Allergy Asthma Immunol. 2011;106:533-7.e2.
- Parmiani S, Fernández Távora L, Moreno C, Guardia P, Rico P. Clustered schedules in allergen-specific immunotherapy. Allergol Immunopathol (Madr). 2002;30:283-91.