SPECIAL ARTICLE

The Interleukin Network in the Immunopathogenesis of Oral Diseases

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A remarkable breakthrough in the understanding of both cellular and molecular mechanisms of cell to cell communication has been documented in the past two decades. It is now clear that signals provided by accessory molecules such as adhesion molecules and cytokines are necessary for cell activation.^{1,2} Of these cytokines, interleukins produced by both lymphoid and non-lymphoid cells play a crucial role in the immune response.

By the end of 1993, at least 13 interleukins had been discovered. The biochemical and biological properties of the interleukins are beyond the scope of this paper and readers may refer to extensive reviews elsewhere,²⁻⁸ The functions of interleukins depend upon their specific receptors on the cell surface; however, a precise mechanism of signal transduction resulted from interleukin-receptor binding remains to be elucidated. The molecular and functional characteristics of these receptors have been reviewed recently.9

The exact pathogenesis of certain oral diseases such as chronic inflammatory periodontal disease

SUMMARY Interleukins produced by both lymphoid and non-lymphoid cells play a crucial role in the immune response. This paper discusses the possible interleukin network in the immunopathogenesis of some oral diseases. In chronic inflammatory periodontal diseases and periapical inflammation, interleukin's such as IL-1 and IL-6 may be responsible in tissue destruction. High levels of IL-12 but not IL-4 and IL-10 may reduce the course of candidal infection. The progression of HIV infection has been associated with the regulation of distinct cytokines; thus, the pathogenesis of Kaposi's sarcoma may be regulated by IL-6. In autoimmune-associated oral diseases such as lichen planus, the role of Langerhans cells in presenting autoantigens may parallel with increased levels of IL-6. It seems, therefore, that the course of these diseases is regulated by these polypeptides which may in turn modulate the disease severity. However, whether altered levels of interleukins in certain oral disorders can be used as a diagnostic marker requires further investigation.

and lichen planus remains unclear. Yet, accumulating evidence suggests that the progression of these diseases are under the control of immune response; therefore, interleukins produced by activated immunocompetent cells would obviously play a significant role in both diseases, although its exact mechanism is largely unknown.^{10,11} The aim of this paper is to provide some insights into the possible role of the interleukins in some oral soft tissue disorders.

Chronic inflammatory periodontal diseases (CIPD)

The precise immunopathogenesis of CIPD is still enigma. It appears that all periodontopathic bacteria-activated immunocompetent cells are involved in the course of this disease.^{11,12} Thus, the number of T cells in the initial or stable lesion are higher than that of B cells, whereas the progressive or

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Correspondence : W. Sosroseno, Department of Dental Public Health, Faculty of Dentistry, Gadjah Mada University, Yogyakarta, Indonesia. advanced lesion is dominated by B cells; therefore, the former and latter lesions can also be termed the T and B cell lesions, respectively.¹¹ The interconversion of these lesional stages remains however questionable. The type and nature of periodontopathic bacteria, type of antigen presenting cells (APC) and the cytokine network may determine the nature and course of CIPD.¹¹⁻¹³

Altered levels of IL-1, IL-2, IL-4, IL-5, IL-6 and IL-8 have all been implicated in the course of CIPD,¹²⁻¹⁴ High levels of IL-1 released by periodontopathic bacteria-stimulated human mononuclear cells¹⁵ are detected in gingival crevicular fluid (GCF) and gingival tissues from periodontal patients^{16,17} suggesting that this cytokine plays a significant role in CIPD, perhaps, by mediating both soft and bone tissue destruction.13,14 Likewise, elevated serum but reduced gingival IL-2 levels in these patients have also been observed.^{18,19} The extrapolation of these findings in the immunopathogenesis of CIPD is however unknown. One may assume that lack of gingival diseased sitederived IL-2 production suggests that suppressed local T (Th1?) cell activation occurs, particularly in the progressive lesion. This speculation remains, however, to be further investigated.

In periodontal patients, elevated levels of IL-5 and IL-6 released by diseased gingival have also been detected. 15, 19-21 Both cytokines are extremely important in both T cell activation and terminal differentiation of activated B cells, 22,23 Therefore, it seems possible that these interleukins play a pivotal role in the progressive lesion, by activating Th2 cells which in turn suppress Th1 cells and help B cell activation (Sosroseno W and Herminajeng E, submitted). Alternatively, since IL-6 has been shown to induce HLA-DR expression on the gingival Langerhans cells,²⁴ increased levels of IL-6 in the inflamed gingival tissues may enhance the antigen presentation function of these Langerhans cells which in turn modulate the local immune response. This cytokine may also have bone resorbing activity, since antibody to IL-6 reduced bone resorption in ovariectomized mice.²⁵ The ability of this cytokine in periodontal bone destruction in CIPD can not, therefore, be ruled out and needs to be further investigated.¹¹

Perhaps one of the most intriguing questions is the role of IL-8 in CIPD. In the presence of IL-1, gingival fibroblasts are able to produce IL-8, suggesting that this cytokine acts as a chemotactic factor for neutrophils during the course of CIPD.²⁶ It is also possible that IL-8 may recruit T cells at the gingival inflamed site, since this cytokine is also chemotactic for T cells.²⁷ This question needs to be further investigated.

Periapical inflammation

Periapical inflammation is also known as an inflammatory dental pulp reaction induced by oral bacteria such as Porphyromonas endodentalis. The progression of this inflammation may be under the control of the immune network.28 Following bacterial invasion and tissue responses, it has been speculated that the activated cells-derived IL-1 induces periapical bone destruction.²⁹ In an animal model, extracts of periapical tissues derived from exposed dental pulp contained polypeptides, including IL-1 and TNF-alpha.³⁰ In this study the induction of fetal rat long bone resorption mediated by these extracts could be suppressed in the presence of anti-IL-1 alpha antibodies, suggesting that IL-1 production in the inflamed dental pulp may be responsible for periapical bone damage. It is however unclear whether other interleukins would be involved in the course of this

lesion; it may be that IL-6 produced by various cells in this site participates in the tissue destruction by activating osteoclasts as speculated in the immunopathogenesis of CIPD.¹¹

Candidiasis

Candidiasis caused by Candida albicans is perhaps one of the best example of oral fungal diseases. It appears that antigen-specific T cell response is dominant during the course of C.albicans-induced immunological reaction; thus, depletion of CD4 cells in vivo inhibits C.albicans-induced DTH reaction and enhances the disease severity. 31,32 More recent studies revealed that, dependent upon distinct C.albicansisolated antigens used, both IL-2 and IFN-gamma-producing Th1 cells are activated in genetically resistant mice such as BALB/cCr mice, whereas IL-4 and IL-6-producing Th2 cells are associated with genetically susceptible mice such as DBA/2 mice.^{33,34} suggesting that the progression of candidal infection may be associated with the function of these distinct cytokines (Fig. 1). The fact that Th1 cell-derived IFN-gamma enhances the antifungal activity of macrophages, perhaps by increasing nitric oxide production especially in a murine model^{35,36} supports a pivotal role of cytokines in candidiasis. However, whether elevated levels of IL-4 reflect a late protection or increased susceptibility remains to be elucidated. It appears in this respect that the latter speculation may be the case, since the course of the disease could be inhibited and both DTH responsiveness and long lasting protection were observed in IL-4-neutralized murine candidiasis.³⁷ Blocking IL-4 activity by injection of recombinant IL-4 receptor resulted in depressed candidal infection in genetically susceptible mice.³⁸ Likewise, the induction of IL-4 mRNA expression following a single injection of



monoclonal anti-IFN-gamma antibodies parallels the loss of DTH response and specific protection in murine candidiasis.³⁹

The exact mechanism by which Th1 to Th2 cell shift is associated with the progression of candidiasis remains to be elucidated. Romani and colleagues have shown that IL-12 mRNA expression in macrophages of genetically resistant but not susceptible mice is detected.40 In this study, IL-12 was required to develop C.albicans specific-Th1 cells, suggesting that Candidaactivated macrophages produce IL-12 to develop antigen-specific Th1 cells. It is also possible that certain Candida-derived superantigens such as heat shock protein 90kD (HSP 90) may suppress the development of Th2 cells and induce Th1 cells which in turn enhance resistance to candidal infection.41,42 In sharp contrast, addition of IL-4 and IL-10 downregulates the production of nitric oxide and murine macrophage antifungal activity to *C.albicans in vitro.*³⁶ *In vivo*, injection of antibody to IL-10 enhanced nitric oxide and IFN-gamma productions and protected genetically susceptible mice to candidiasis,⁴³ suggesting that IL-10 plays a major role in suppressed Th1 cell development and thus, enhances susceptibility to candidal infection.

The extrapolation of these findings to humans is speculative, however, since unlike the murine model, IL-10 is produced both by human Th1 and Th2 cells.⁴⁴ Notwithstanding, the cytokine profiles in *C.albicans*-induced immune response which may determine the susceptibility of the infected individuals certainly provide a new insight into the immunopathogenesis of candidiasis and therapeutic bimodality of cytokines in this pathological entity.

AIDS-associated oral diseases

The precise immunopathogenesis of AIDS (acquired immunodeficiency syndrome) caused by HIV-1, a retrovirus, is still an enigma. Candidiasis, hairy leukoplakia and Kaposi's sarcoma are among the most common oral disoders seen in the HIV-infected patients.⁴⁵

IL-1, IL-2, IL-6, TNF-α and IFN-gamma have all been implicated in the immunopathogenesis of AIDS.^{46,47} For example, despite a T cell defect, hyperactivated B cells have been suggested to be due to overproduction of IL-6 in HIVinfected patients.22,23 Increased levels of IL-4 are also detected and accompanied by a lack of IL-2 production in the progression of AIDS; however, the levels of IL-2 but not IL-4 are dominant in asymptomatic seropositive patients.⁴⁸ Based upon these findings, it is believed that during the course of HIV infection, Th1 cell-derived IL-2 provides protection, whereas Th2 cell-derived IL-4 and IL-6 would induce progressive or symptomatic disease.49 It may therefore be that lack of IL-2 and IFN-gamma production reduces CD8 cell activities to lyse the HIV-infected cells in the symptomatic stage.49 In this hypothesis, the switch may be due to the production of Th2 cell-derived IL-10 which in turn suppresses Th1 cell activation.

Yet, this hypothesis remains arguable. Unlike the murine system, both human Th1 and Th2 cell clones produce IL-10 which in turn inhibits their proliferation and cytokine production.⁴⁴ Should the Th1 and Th2 cell shift occur, it may require explanations other than simple mediation by IL-10 *per se*. One possibility is that HIV-infected APC induces Th1 cell anergy, thereby preferentially expanding Th2 cells.⁵⁰ In this respect, lack of APC-derived IL-1 and/or IL-12 may play a major role in inducing IL-2 and IFNgamma-producing Th1 cell anergy and vice versa.^{51,52} Whether this shift which determines the course of AIDS is orchestrated by these pathways remains speculative.

The most recent studies have shown clearly that the progression of HIV infection is not merely associated with Th1 to Th2 cell shift.53,54 In these studies, peripheral blood and lymph node cell-derived IFNgamma and IL-10, but not IL-2 and IL-4 could be detected in both HIV-seronegative and -seropositive subjects. Following HIV stimulation in vitro, CD4 cells from these subjects produced IFN-gamma, IL-4 and IL-10, cytokines belonging to Th0 cells. These results suggested that HIV infection did not induce a Th1 to Th2 cell shift, rather it would stimulate the development of Th0 cells.

Although the hypothesis based upon the HIV-induced cytokine profiles remains to be clarified further, progressive AIDS-associated oral diseases may be predicted from this point of view. With respect to Kaposi's sarcoma, spindlelike cells isolated from this disease produce cytokines such as IL-6 and IL-1, but not IL-2 and IFNgamma, suggesting that the progressive stage of this disease is associated with the cytokine profile.⁵⁵ This idea needs, however, to be investigated further.

Autoimmune-associated oral diseases

Autoimmune response can perhaps be considered as a failure of the immune system to discriminate between self-and non self-antigens. In the oral cavity, various soft tissue lesions have been associated with autoimmune diseases such as lichen planus,¹⁰ Sjörgren's syndrome,⁵⁶ pemphigoid,⁵⁷ Behçet syndrome,⁵⁸ and systemic lupus erythematosus.⁵⁹ With the exception of pemphigoid, the exact autoantigens, if these are the cause of autoimmune diseases, are largely unknown and their clinical appearances in the oral cavity are likely to be a manifestation of systemic rather than local immune disorders.

The detailed mechanisms of autoimmunity such as failure of deleted self-reactive T and B cells^{60,61} and of gene-induced apoptosis,62 as well as downregulated CD8 cell functions⁶³ and superantigens-induced immune response⁶⁴ are beyond of the scope of this paper. Of interest, cytokines released by the autoantigens-activated immunocompetent cells are a crucial factor in expanding autoreactive T and B cells as well as generating tissue destruction which exposes the hidden autoantigens.65,66 Thus, in insulin-dependent diabetes mellitus, IL-1 produced by activated macrophages could induce islet Beta cell damage which results in releasing its autoantigens.⁶⁷ Likewise, increased levels of IL-4 may maintain B cell hyperactivation seen in SLE patients.68 Joint destruction seen in rheumatoid arthritis may be ameliorated by TGF- β which may inhibit IL-1 production.69

Lack of conclusive evidences concerning altered interleukin levels in autoimmune-related oral diseases leads to a difficulty in determining the role of these biologically active polypeptides. In lichen planus, it has been hypothesized that damaged keratinocytes would release IL-1 and IL-6 to stimulate Langerhans cell functions.¹⁰ The IL-6 levels were significantly elevated in active disease and decreased following remission, suggesting that it plays a significant role in lichen planus, perhaps by enhancement of MHC class II expression on Langerhans cells.^{24,27} In Sjorgren's syndrome, salivary gland biopsies obtained from the patients and cultured in vitro produced IL-2 and another B cell growth factor, perhaps IL-10.71,72 It was thought that these

cytokines would maintain autoreactive T and B cell growth in this syndrome.⁵⁶ However, since IgG1 antibodies of Sjörgren's syndrome patients are selectively produced in high levels,⁷³ an excessive production of IL-4 may also occur.

Oral neoplasia

The development of cancer cells requires a complex network including growth factors. The roles of transforming growth factoralpha and -beta as well as of epidermal growth factor in cancer cells are beyond the scope of this paper. Certain interleukins appear to be important factors in cancer growth. Lung cancer cells spontaneously produce significant levels of IL-874,75; however, the role of this cytokine in tumor growth remains unclear. In IL-9 transgenic mice, 7% of these mice developed thymic lymphomas.⁷⁶ In this study, expression of IL-9 was required for optimal tumor growth, suggesting that overproduction of IL-9 may induce the development of thymic tumor cells. These results are also confirmed by the fact that human anaplastic large cell lymphomas require IL-9 in an autocrine fashion.77 Possibly cancer cell-derived IL-10 is one of the examples of ways by which these cells evade the host immune defences. Human B cell lymphomas, lung cancer and melanoma are capable of producing IL-10 which suppresses tumoricidal functions of monocytes and macrophages.78-81 Nabioullin and colleagues have also shown that a cocktail of IL-10 and IL-14 is much more potent as an inhibitory factor to these cells than either IL-10 or IL-4 alone.81 Melanomaderived IL-10 inhibits the production of TNF-alpha, IFN-gamma, and IL-2 and suppresses mixed lymphocyte reactions.82

The concept of the immunosurveillance carried out by NK cells, T cells and macrophages is important in tumor rejection.⁸³⁻⁸⁵ In fact, activated cells-derived cytokines which have anti-tumor activities are also crucial effectors in rejecting tumor growth. Dependent upon the tumor origin, expressed IL-2 and IFN-gamma mRNA were observed in basal cell carcinoma biopsies, whereas IL-4, IL-5 and IL-10 mRNA were highly expressed in seborrheic keratosis, a benign growth of the epidermis.⁸⁶ It seems therefore that along with other cytokines such as TNF-alpha, IL-2 and IFNgamma play an important role in tumor rejection, despite the fact that tumor-activated IL-4 and IL-6-producing T (Th2) cells can inhibit tumor growth.⁸⁷ Perhaps IL-2 and IFN-gamma are needed to recruit and activate cytotoxic CD8 T cells and NK cells as well as macrophages. IL-6 may augment the host anti-tumor defences by, for example, increased antibody production and elevated T and B cell activation; on one hand, IL-4 could induce the LAK (lymphokine-activated killer) cells activity.88,89 It should be noticed that the anti-tumor function of IL-6 is only limited on the early but not advanced stage of the tumor growth, 90 implying that IL-6induced the host's defences only act on the initial tumor growth, whilst IL-2 and IFN-gamma-activated defences function in the advanced disease stage. It has been revealed that IL-7 is able to act as an antitumor agent, by inducing monocyte and macrophage-derived IL-1 alpha, IL-1 beta and TNF-alpha production.91

Surprisingly, evidence for a role of interleukins in the development of oral neoplasia is lacking. The work of Gangal and Tatake⁹² has shown that impaired IL-2 production is observed in patients with oral leukoplakia; however, normal production of this cytokine is seen in untreated oral cancers. It is of interest that reduced cytotoxic functions of NK cells and LAK cells could be restored in addition of IFN-gamma and IL-2,⁹² implying

that in oral cancers, altered IFNgamma rather than IL-2 production occurs. Yet, it is unclear whether IL-4, IL-6 and IL-7 would contribute to inhibition of the development of oral neoplasia.

Conclusion

The signals provided by the interleukins in the immune response prove to be prerequisites. Thus, transient levels of interleukins would determine the efficacy of the immune response against various antigens. In microbial antigen-induced oral diseases, the levels of IL-1, IL-2, IL-4, IL-5 and IL-6 would affect the course of the diseases and individual susceptibility. Likewise. these biologically active polypeptides may cause deterioration in the progression of autoimmune diseases, determining some of the clinical features in the oral cavity. In AIDS-associated oral diseases, the interleukin profiles seen in HIVinfected humans may be parallel with the course of these diseases. Furthermore, although altered IL-2 levels could be observed in oral cancers, it is not clear whether it reflects suppressed host immune defences and whether other cytokines are involved in the cancer growth and regression.

Despite the fact that production of some interleukins is impaired in some oral diseases, it is still uncertain whether they can be utilized as diagnostic markers. In this respect, the levels of other cytokines, such as TNF-alpha and TGF-beta which regulate the production of interleukins during the course of oral diseases need to be considered.

REFERENCES

- Springer TA. Adhesion receptors of the immune system. Nature 1990; 346 : 425-34.
- 2. Arai KI, Lee F, Miyajima A, Miyatake S, Arai N, Yokota T. Cytokines : Coor-

dinators of immune and inflammatory responses. Ann Rev Biochem 1990; 59 : 783-836.

- Mukaida N, Harada A, Yasumoto K, Matsushima K. Properties of proinflammatory cell type-specific leukocyte chemotactic cytokines, interleukin 8 (1L-8) and monocyte chemotactic and activating factor (MCAF). Microbiol 1mmunol 1992; 36 : 773-89.
- Renauld J-P, Houssiau F, Druez C, Uyttenhove C, Vink A, Van Snick J. Interleukin-9. Int Rev Exp Pathol 1993; 34A: 99-109.
- Rennick D, Berg D, Holland G. Interleukin 10 : an overview. Progr Growth Factor Res 1992; 4 : 207-27.
- Yang Y-C, Yin T. Interleukin-11 and its receptor. Biofactors 1992; 4: 15-21.
- Trinchieri G. Interleukin-12 and its role in the generation of T_H1 cells. Immunol Today 1993; 14: 335-7.
- Zurawski G, de Vries JE. Interleukin 13, an interleukin 4-like cytokine that acts on monocytes and B cells, but not on T cells. Immunol Today 1994; 15 : 19-26.
- Foxwell B, Barrett K. Introduction to cytokine receptors : structure and signal transduction. Int Rev Exp Pathol 1993; 34B : 105-21.
- Walsh LJ, Savage NW, Ishii T, Seymour GJ. Immunopathogenesis of oral lichen planus. J Oral Pathol Med 1990; 19: 389-96.
- Seymour GJ, Gemmell E, Reinhardt RA, Eastcott J, Taubman MA. Immunopathogenesis of chronic inflammatory periodontal disease : cellular and molecular mechisms. J Periodont Res 1993; 28 : 478-86.
- Genco RJ. Host responses in periodontal diseases : current concepts. J Periodontol 1993; 63 : 338-5.
- Page R. The role of inflammatory mediators in the pathogenesis of periodontal disease. J Periodont Res 1991; 26 : 230-42.
- Kjedsen M, Holmstrup P, Bendtzen K. Marginal periodontitis and cytokines : A review of the literature. J Periodontol 1993; 64 : 1013-22.
- Gemmell E, Seymour GJ. Interleukin 1, interleukin 6 and transforming growth factor-B production by human gingival m tonuclear cells following stimulation

with Porphyromonas gingivalis and Fusobacterium nucleatum. J Periodont Res 1993; 28 : 122-9.

- Kinane DF, Winstanley FP, Adonogianaki E, Moughal NA. Bioassay of interleukin l (IL-1) in human gingival crevicular fluid during experimental gingivitis. Arch Oral Biol 1992; 37 : 153-6.
- Stashenko P, Fujiyoshi P, Obernesser MS. Prostak L, Haffajee AD, Socransky SS. Levels of interleukin 1β in tissue from sites of active periodontal disease. J Clin Periodontol 1991; 19 : 548-54.
- McFarlane CG, Meikle MC. Interleukin-2, interleukin-2 receptor and interleukin-4 levels are elevated in the sera of patients with periodontal disease. J Periodont Res 1991; 26 : 402-8.
- Fujihashi K, Kono Y, Beagley KW, Yamamoto M, McGhee JR, Mestecky J, Kiyono H. Cytokines and periodontal disease : immunopathological role of interleukins for B cell responses in chronic inflamed gingival tissue. J Perio dontol 1993; 64 : 400-6.
- Geivelis M, Turner DW, Pederson ED, Lambert BL. Measurements of interleukin-6 in gingival crevicular fluid from adults with destructive periodontal disease. J Periodontol 1993; 64 : 980-3.
- Takahashi K, Takashiba S, Nagai A, Takigawa M, Myoukai F, Kurihara H, Murayama Y. Assessment of interleukin-6 in the pathogenesis of periodontal disease. J Periodontol 1994; 65: 147-53.
- Sosroseno W, Herminajeng E. Interleukin 6 (IL-6) : the biochemistry and its role in B and T cell development. Med J Univ Indon (In press).
- Hirano T, Kishimoto T. (eds). Molecular biology and immunology of interleukin-6. Res Immunol 1992; 143 : 723-83.
- Ishii T, Walsh LJ, Seymour GJ, Powell RN. Modulation of Langerhans cell surface antigen expression by recombinant cytokines. J Oral Pathol Med 1990; 19: 355-9.
- Jilka RL, Hangoc G, Girasole G, et al. Increased osteoclast development after estrogen loss : mediation by interleukin-6. Science 1992; 257 : 88-91.
- 26. Odake H, Koizumi F, Hatakeyama S,

Furuta I, Nakagawa H. Production of cytokines belonging to the interleukin-8 family by human gingival fibroblasts stimulated with interleukin-1 β in culture. Exp Mol Pathol 1993; 58 : 14-24.

- Larsen CG, Anderson AO, Appella E, Oppenheim JJ, Matsushima K. The neutrophil-activating protein (NAP-1) is also chemotactic for T lymphocytes. Science 1989; 243 : 1464-6.
- Jontell M, Bergenholtz G. Accessory cells in the immune defence of the dental pulp. Proc Finn Dent Soc 1992; 28 (suppl. 1) : 345-55.
- 29. Stashenko P. The role of immune cytokines in the pathogenesis of periapical lesions. Endod Dent Traumatol 1990; 6: 89-96.
- Wang C-Y, Stashenko P.The role of interleukin-1α in the pathogenesis of periapical bone destruction in a rat model system. Oral Microbiol Immunol 1993; 8 : 50-6.
- Cantorna TT, Balish E. Role of CD4⁺ lymphocytes in resistance to mucosal candidiasis. Infect Immun 1991; 59 : 2447-55.
- 32. Romani L, Mencacci A, Cenci E, et al. Course of primary candidiasis in T-celldepleted mice infected with attenuated variant cells. J Infect Dis 1992; 166 : 1384-92.
- 33. Romani L, Mocci S, Bietta C, Lanfaloni L, Puccetti P, Bistoni F. Th1 and Th2 cytokine secretions patterns in murine candidiasis : association of Th1 responses with acquired resistance. Infect Immun 1991; 59 : 4647-54.
- 34. Romani L, Mencacci A, Cenci E, et al. CD4 + cell subset expression in murine candidiasis. The responses correlate directly with genetically determined susceptibility or vaccine-induced resistance. J Immunol 1993; 150 : 925-31.
- Redmond HP, Shou J, Gallagher HJ, Kelly CJ, Daly JM. Macrophagedependent candidacidal mechanisms in the murine system. J Immunol 1993; 150: 3427-33.
- 36. Cenci E, Romani L, Spaccapelo R, et al. Interleukin-4 and interleukin-10 inhibit nitric oxide-dependent macrophage killing of Candida albicans. Eur J Immunol 1993; 23 : 1034-8.
- 37. Romani L, Mencacci A, Grohmann U,

et al. Neutralizing antibody to interleukin 4 induces systemic protection and T helper type 1-associated immunity in murine candidiasis. J Exp Med 1992; 176 : 19-25.

- Puccetti P, Mencacci A, Cenci E, et al. Cure of murine candidiasis by recombinant soluble interleukin-4 receptor. J Infect Dis 1994; 169 : 1325-31.
- Romani L, Cenci E, Mencacci A, et al. Gamma interferon modifies CD4⁺ subset expression in murine candidiasis. Infect Immun 1992; 60 : 4950-2.
- Romani L, Mencacci A, Tonnetti L, et al. Interleukin-12 but not interferon-% production correlates with induction of T helper type-1 phenotype in murine candidiasis. Eur J Immunol 1994; 24 : 909-15.
- Matthews RC. Candida albicans HSP 90 : link between protective and autoimmunity. J Med Microbiol 1992; 36 : 367-70.
- Romani L, Puccetti P, Mencacci A, et al. Tolerance to staphylococcal enterotoxin B initiates Th1 cell differentiation in mice infected with Candida albicans. Infect Immun 1994; 62: 4047-53.
- Romani L, Puccetti P, Mencacci A, et al. Neutralization of 1L-10 upregulates nitric oxide production and protect susceptible mice from challenge with Candida albicans. J Immunol 1994; 152 : 3514-21.
- 44. Del Prete G, De Carli M, Almerigogna F, Giudizi MG, Biagiotti R, Rogmanani S. Human IL-10 is produced by both type 1 helper (Th1) and type 2 helper (Th2) T cell clones and inhibits their antigen-specific proliferation and cytokine production. J Immunol 1993; 150 : 353-60.
- 45. Verdon R, Bouchaud O, Rene E. Oral and aesophageal diseases in human immunodeficiency virus-infected patients. Eur J Gastroenterol Hepatol 1992; 4 : 396-403.
- Rosenberg ZF, Fauci AS. Immunopathogenic mechanisms of HIV infection : cytokine induction of HIV. Immunol Today 1990; 11 : 176-80.
- 47. Doweiko JP. Hematologic aspects of HIV infection. AIDS 1993; 7 : 753-7.
- Clerici M, Hakim FT, Venzon DJ, et al. Changes in interleukin-2 and interleukin-4 production in asymp-

tomatic, human immunodeficiency virus-seropositive individuals. J Clin Invest 1993; 91 : 759-65.

- 49. Clerici M, Shearer GM, A $T_H I$ $T_H 2$ switch is a critical step in the etiology of HIV infection. Immunol Today 1993; 14 : 107-11.
- Meyaard L, Schuitemaker H, Miedema F, T-cell dysfunction in HIV infection : anergy due to defective antigen-presenting cell function? Immunol Today 1993; 14 : 161-4.
- Schmitz J, Assenmacher M, Radbruch A. Regulation of T helper cell cytokine expression : functional dichotomy of antigen-presenting cells. Eur J Immunol 1993; 23 : 191-9.
- 52. Manetti R, Parronchi P, Giudizi MG, et al. Natural killer cell stimulatory factor (interleukin 12 [1L-12] induces T helper type 1 (Th1)-specific immune responses and inhibits the development of 1L-4-producing Th cells. J Exp Med 1993; 177 : 1199-204.
- 53. Maggi E, Mazzetti M, Ravina A, et al. Ability of HIV to promote a $T_{H}1$ to $T_{H}0$ shift and to replicate preferentially in $T_{H}2$ and $T_{H}0$ cells. Science 1994; 265 : 244-8.
- Graziosi C, Pantaleo G, Gantt KR, et al. Lack of evidence for the dichotomy of T_H2 and T_H2 predominance in HIV-infected individuals. Science 1994; 265 : 248-52.
- 55. Ensoli B, Barillari G, Gallo RC. Cytokines and growth factors in the pathogenesis of AIDS-associated Kaposi's sarcoma. Immunol Rev 1992; 127: 147-55.
- Fox RI, Kang H-1. Pathogenesis of Sjögren's syndrome. Rheum Dis Clin North Am 1992; 18: 517-38.
- Anhalt GJ. Pemphigoid : Bullous and cicatricial. Dermatol Clin 1990; 8 : 701-14.
- Powell RJ, Dunstan S. Immunopathology of Behçet's disease. Postgrad Med J 1991; 67: 503-5.
- 59. Johnson R, Holdahl R. Infiltrating mononuclear cells in salivary glands and kidneys in autoimmune MRL/ Mp-2pr/1pr mice express 1L-2 receptor and produce interferon-γ. J Oral Pathol Med 1990; 19: 330-4.
- 60. Lo D. T cell-tolerance. Curr Opin Immunol 1992; 4 : 711-5.

- 61. Goodnow CC. B-cell tolerance. Curr Opin Immunol 1992; 4 : 703-10.
- Cohen PL, Eisenberg RA. The *lpr* and *gld* genes in systemic autoimmunity : life and death in the *Fas* lane. Immunol Today 1992; 13 : 427-28.
- 63. Tomer Y, Shoenfeld Y. The significance of T suppressor cells in the development of autoimmunity. J Autoimmun 1989; 2 : 739-58.
- 64. Acha-Orbe H. Bacterial and viral superantigens : role in autoimmunity. Ann Rheum Dis 1993; 52 : S6-S16.
- Brennan FM, Feldman M. Cytokines in autoimmunity. Curr Opin Immunol 1992; 4: 754-9.
- Opdenaker G, Van Damme J. Cytokineregulated proteases in autoimmune diseases. Immunol Today 1994; 15 : 103-6.
- Molvig J. A model of the pathogenesis of insulin-dependent diabetes mellitus. Danish Med Bull 1992; 39: 509-41.
- Linker-Israeli M. Cytokine abnormalities in human lupus. Clin Immunol Immunopathol 1992; 63: 10-2.
- Sosroseno W, Herminajeng W. The immunoregulatory role of transforming growth factor beta (TGF-B) : A review. Br J Biomed Sci (In press).
- 70. Yamamoto T. Yoneda K, Ueta E, Hirota J, Osaki T. Serum cytokine levels in patients with oral mucous membrane disorders. J Oral Pathol Med 1991; 20: 275-9.
- Fox R1, Theofiloppoulos AN, Altman A. Production of interleukin 2 (1L-2) by salivary gland lymphocytes in Sjörgren's syndrome. Detection of reactive cells by using antibody directed to synthetic peptides of 1L-2. J Immunol 1985; 135 : 3109-15.
- 72. Takei M, Dang H, Dauphinee MJ, Talal N. Autostimulatory growth factors produced by Sjörgren's syndrome B cell lines. Clin Immunol Immunopathol 1989; 53 : 3-35.
- 73. Hay EM, Freemont AJ, Kay RA, Berstein RM, Holt PJL, Pumphrey RSH. Selective polyclonal increase of immunoglobulin G1 subclass: a link with Sjörgren's syndrome. Ann Rheum Dis 1990; 49: 373-7.
- 74. Mizuno K, Sone S, Orino E, et al. Spontaneous production of interleukin-8 by human lung cancer cells and its

augmentation by tumor necrosis factor alpha and interleukin-1 at protein and mRNA levels. Oncology 1994; 51 : 467-71.

- Smith DR, Plverini PJ, Kunkel SL, et al. Inhibition of interleukin 8 attenuates angiogenesis in bronchogenic carcinoma. J Exp Med 1994; 179 : 1409-15.
- Renauld JC, Van der Lugt N, Vink A, et al. Thymic lymphomas in interleukin 9 transgenic mice. Oncogene 1994; 9 : 1327-32.
- Hsu SM, Waldron JW, Jr Hsu PL, Hough AJ Jr. Cytokines in malignant lymphomas : review and prospective evaluation. Human Pathol 1993; 24 : 1040-57.
- Stewart JP, Behm FG, Arrand JR, Rooney CM. Differential expression of viral and human interleukin-10 (IL-10) by primary B cell tumors and B cell lines. Virology 1994; 200 : 724-32.
- Smith DR, Kunkel SL, Burdick MD, et al. Production of interleukin-10 by human bronchogenic carcinoma. Am J Pathol 1994; 145 : 18-25.
- 80. Luscher U, Filguera L, Juretic A, et al. The pattern of cytokine gene expression in freshly excised human metastatic melanoma suggests a state of reversible anergy of tumor-infiltrating lymphocytes. Int J Cancer 1994; 15 : 612-9.
- Nabioullin R, Sone S, Mizuno K, et al. Interleukin-10 is a potent inhibitor of tumor cytotoxicity by human monocytes and alveolar macrophages. J Leukocyte Biol 1994; 55 : 437-42.
- Chen Q, Daniel V, Maher DW, Hersey P. Production of 1L-10 by melanoma cells : examination of its role in immunosuppression mediated by melanoma. Int J Cancer 1994; 56 : 755-60.
- Ioannides CG, Whiteside TL. T cell recognition of human tumors : implications for molecular immunotherapy of cancer. Clin Immunol Immunopathol 1993; 66 : 91-106.
- Mantovani A, Bottazzi B, Colotta F, Sozzani S, Ruco L. The origin and function of tumor-associated macrophages. Immunol Today 1992; 13: 265-70.
- Whiteside TL, Herberman RB. The role of natural killer cells in human disease. Clin Immunol Immunopathol 1989; 53 : 1-23.
- 86. Yamamura M, Modlin RJ, Ohmen JD,

Moy RL. Local expression of antiinflammatory cytokines in cancer. J Clin Invest 1993; 91 : 1005-10.

- 87. Lauritzsen GF, Weiss S, Bogen B. Anti-tumor activity of idiotypespecific, MHC-restricted Th1 and Th2 clones in vitro and in vivo. Scand J Immunol 1993; 37 : 77-85.
- 88. Revel M. Growth regulatory function of 1L-6 and antitumor effects. Res Immunol 1992; 143 : 769-73.
- 89. Mule JJ, Smith CA, Rosenberg SA. 91. Alderson MR, Tough TW. Ziegler SF. Interleukin-4 (B-cell stimulatory factor-1) can mediate the induction of LAK activity directed against fresh tumor cells. J Exp Med 1987; 166 : 792-7.
- 90. Kerbel, RB. Expression of multicytokine resistance and multi-growth factor independence in advanced stage metastatic cancer. Am J Pathol 1992; 141 : 519-25.
- Grabstein KH. Interleukin 7 induces cytokine secretion and tumoricidal activity by human peripheral blood monocytes. J Exp Med 1991; 173 : 923-30.
- 92. Gangal SG, Tatake RJ. Immune status in head and neck cancer with special reference to cancer of the oral cavity. In : Johnson NW, ed, Oral cancer. Cambridge : Cambridge Univ. Press., 1991 : 114-54.