SPECIAL ARTICLE

Characterzation of Immunological Memory Cells. A Minireview.

Molvibha Vongsakul

In general parlance, memory means the ability to store and to recall something. The meaning is the same for immunological memory. Moreover, in immunology we usually consider that memory functions by shortening the time required for the positive detection of antigens and especially for the rapid increase in production of specific high affinity antibodies. What are the differences among immunological memory cells and other immunological cells? Do we have both B and T memory cells. How long can immunological memory cells survive? How do they circulate? Where are they located? How can we detect or demonstrate them? These are some interesting questions that this article will try to address. Many studies have been performed and many hypotheses have been proposed, however, they are somewhat inconclusive. This minireview considers some aspects of immunological memory cells according to (1) lineage, (2) longevity and recirculation pattern, (3) phenotypic markers expression, and (4) detection methods.

Lineage of immunological memory B and T cells

The existence of memory B cells

SUMMARY : This paper reviews the information concerning immunological memory T and B cells. It shows that the existence of different or a single lineage of memory and naive cells is still a question. The recirculation pattern of memory cells is different from naive cells. A unique recirculation for memory T or B cells is suggested while the bcl2 gene expression possibly plays role in the longevity of memory cells. Various phenotypic markers are demonstrated only on memory or on naive cells; however, a reliable and convenient method for the detection of memory cells still needs to be explored.

has been well recognised for decades, contrary to memory T cells which are comprised of various subpopulations. Therefore, greater controversy exists concerning the role of memory T cells in the immune response than about the role of memory B cells. The first criterion which indicates the presence of memory B cells is the shortened time required for positive detection of specific antibodies with high or increasing affinity for a given antigen. The function of memory T cells is more complicated since they encompass subpopulations of T cells and they cause no change in immunoglobulin isotype as observed with memory B cells. Nevertheless, memory T cells do exhibit specific activities when tested in vitro in comparison to naive T cells¹ and there is evidence for an increase in frequency of precursor memory T cells associated with certain viral infections.²

There are at least two ideas concerning memory B-cell development. The first proposes that there is only one lineage for B cells that leads to either naive cells or memory cells. According to this theory, some cells differentiate and mature into antibody producing cells upon interaction with specific antigens while others are arrested in their differentiation at a certain stage and become quiescent until reexposed to the same antigen, after which they resume differentiation into antibody producing cells.^{3,4} T-cell derived factors may be involved in B cell differentiation. For example, the lack

From the Department of Microbiology. Faculty of Science, Mahidol University Bangkok 10400, Thailand.

of an appropriate T-cell factor at some stage of B cell differentiation may prevent development of antibodyproducing cells. This would suggest the existence or release of qualitatively different suppressive factors from T cells upon antigenic stimulation.⁴ The second theory for B-cell development proposes that there are different lineages of B cells which become antibody producing cells or memory cells. This concept is supported by the existence of a specific, phenotypic marker, J11D, which can be detected at varying levels on B cells by a specific monoclonal antibody. The speculation is that B cells which express high levels of the J11D marker upon stimulation with specific antigen will generate antibody producing cells while B cells which express low levels J11D marker will generate memory cells.⁴⁻⁶ Other evidence suggests that the microenvironment in various parts of lymphoid germinal centers may lead to hypermutation of VH genes with high antibody affinity or to the generation of memory B cells.⁷⁻¹⁰

Information from viral infections and from carrier models demonstrates the existence of both T helper and T suppressor memory cells.^{11, 12} Which T-cell lineage acts as effector/regulator or memory effector/regulator is controversial. Recent studies favor the idea of only one lineage of T cells based on the leukocyte common antigen phenotypic marker CD45 (CD45RA and CD45RO). The work shows that naive T cells contain CD 45RA while memory T cells contain the CD45RO isoform. Whether CD 45 isoforms can be interchangeable is under investigation. At present, some studies suggest that CD45RA-positive T cells can become CD45RO positive cells.4, 6, 13-15 If proven correct this indicates that either CD45RA or CD45RO can be expressed on the same T cell at different activation stages or at different transition stages.

The question of different or single lineage of memory and naive B,

T cells is still a question, The obscure condition of the development of memory and effector T cells, the question of whether effector cells are end cells or revert to memory cells are challenges for the immunologist.¹⁶ To give answers to these questions, specific or unique phenotypic markers are necessary. Elucidation of how early in lymphocyte ontogeny, certain genes are expressed only on either naive or memory cell should give some useful answers.

Longevity and recirculation of memory B and T cells

The exact life span or half-life of lymphocytes (B and T cells) is still an open question. This lack of knowledge is due to limitations in techniques for following specific B or T cells.¹⁷ However, the approximate life span of naive lympocytes is thought to be 3-4 weeks. Memory B or T cells are believed to be longer living cells, having life spans greater than 1 month. There is even a report of a 10 year life span for memory B or T cells¹⁸ and there is a report of a life span as long as 75 years for immunological memory to infectious agents.¹⁷ In fact, there are two opposing ideas concerning the longevity of naive and memory cells. According to one point of view, naive and memory B or T cells have the same half life but the permanence or consistency depends upon restimulation of naive lymphocytes at the specific antigen depot in follicular dendritic cells of the lymphoid organ to allow them to function as memory B or T cells.⁶ According to another point of view, there is a difference in their half lives. This view derives from studies which demonstrate different recirculation patterns associated with the expression of various adhesion molecules on naive and memory B or T cells.¹⁹⁻²² For example, the recirculation of naive lymphocytes is via the high endothelial venule from peripheral blood into lymph nodes, while memory lymphocytes enter lymph nodes via the afferent pathway.

Thus, memory lymphocytes circulate from the peripheral circulation to tissue and then to lymph nodes.²³ There is also evidence for the existence of two types of memory cells, ie circulatory memory cells and non-circulatory memory cells.²⁴

However, the recirculation patterns of naive and memory B, T cells have been shown more clearly to differ according to the expression of an adhesion molecule, L-selectin, as indicated by the observation that naive B, T cells express more Lselectin than memory B cells.²⁵ Nevertheless the recirculation pattern of memory T cells may be different from memory B cells because there is evidence that a carbohydrate molecule termed cutaneous lymphocyte associated antigen (CLA) the ligand for L selectin is expressed only on memory T cells.²⁶

Antiapoptotic function of the bcl2 gene in B and T cell survival has been suggested. For example, it has been shown that upon antigen stimulation in the presence of interleukin-2 some cells die by apoptosis while some survive and function as memory cells.^{27, 28} It is suggested that the product of the bcl2 gene plays a role in lymphocyte survival by influencing apoptosis.²⁹ Although there is no direct evidence to indicate differences in longevity between naive and memory cells there is indirect evidence for a role of the survival gene bcl2 in memory cells to give longer half lives of memory B and T cells than naive cells. Moreover, transgenic mice overproducing bcl2 have been shown to have longer lived memory B cells³⁰ while T cells in homozygous bcl2 deleted mice had a shorter life span than in bcl2 expressed mice.31

Phenotypic markers of memory B and T cells

Phenotypic markers expressed on the surfaces of lymphocytes are important for their circulation and activities. Naive and memory (B or T) cells certainly express surface phenotypic markers and adhesion molecules differently. Some of the markers expressed differently are shown in Table 1. 4, 6, 32-37

However, these are some of the phenotypic markers studied in both humans and mice which some are useful only for mice, for example, Mel14 (Leu 8) and J11D.

How memory B or T cells are detected?

Knowing whether memory B or T cells, and especially their specfic antigens are present is very useful. example, they can help us k whether a test vaccine elicits a poter wanted protection or not and they provide help in diagnosis treatment of diseases. Various in v methods or techniques can be use identify or detect immunolog memory cells. The princples for the in vitro methods are described here.

A. Methods for the detection of memory T cells.

1. Increased proliferation in response to specific antigen challenge in vitro. This can be detected using isotope labelled precursors (nucleotides or amino acids), autoradiography and blast cell staining, for example.³⁸

2. Staining of specific phenotypic markers such as CD45RO, CD44 and IL-2R P75 which are receptors for IL-2. These are the most reliable markers for memory T cells at present. In addition, a recent study has indicated that different adhesion molecule subunit combinations (alpha 4, beta 1) in memory and naive T cells (ie heterogeneous for memory T cells, and homogeneous for naive T cells)³⁶ can also be used. Staining techniques include fluorescence, and immunochemical staining, when results can be obtained using a fluorescence activated cell sorter (FACS).³⁹

3. Detection of cytokines, for example, IL-2 IL-4 and interferon (IFN) production.⁴⁰⁻⁴² In the biological assay, CTLL cell line proliferation can be used for IL-2 detection⁴³

. For	CD 38	-
	CD 39	++
now	CD 44	++
nt or	CD 45 RA	+++
/ can	CD 45 RO	-
and	CD 54	-
vitro	CD 56	
ed to	CD 58	+
gical	CD 69	-
hese	CD 73	+
P	IL-2 R (p 75)	-

MHC II Leu 8 (Mel 14)

J11D

or CT-4S for IL-4 detection. By gel hybridization (mRNA-cDNA dot blot hybridization) specific mRNA cytokines can be detected.44

4. ELISA method. The ELISA plaque assay and ELISPOT assay are techniques for the detection of memory T cells at the single cell level.45,46 These techniques are sensitive and convenient.

B. Methods for the detection of memory B cells.

Two of the above in vitro methods for detection of memory T cells can be used for B cells also. These are:

1. Increased proliferation in response to specific antigen challenge.

2. Staining or detection of certain phenotypic markers such as CD45RO.

Other in vitro methods used only for memory B cells are:

3. Increased quantity or rapidly rising level of IgG after specific antige-

nic challenge. Various methods can be used. Examples are immunoelectrophoresis, radial immunodiffusion and ELISA.

4. Plaque forming assays for detection of antibody producing cells, eg ELISPOT. The advantage of the ELISPOT assay can be shown for the detection of both memory T and memory B cells. This technique can be performed to detect individual cytokine secreting cells for memory T cells or antibody producing cells at the single cell level for memory B cells; quantitation of memory T or B cells is possible.

Since the definite characteristics of naive and memory B, T cells are not completely elucidated, the detection of memory B, T cells using antigen non-specific methods such as detection for the expression of CD45RO, CD44, selectin or the extrusion of P glycoprotein substrate Rhodamine-123⁴⁷ should be performed in parallel as far as possible. The antigen specific

Phenotypic marker	Naive cell	Memory cell
CD 2	++	+++
CD 7	++	±
CD 10	-	+
CD 18/CD 11 a	++	+++
CD 21	++	· ±
CD 23	++	±
CD 25	-	+
CD 26	· •	±
CD 29	+	+++
CD 30	•	++
CD 38	-	++
CD 39	++	-
CD 44	++	+++
CD 45 RA	+++	-
CD 45 RO	-	+++
CD 54	-	+
CD 56	•	+
CD 58	+	++
CD 69	-	±
<u> </u>		

method seems to be reliable, but is time consuming. At the gene level, in parallel detection for certain gene such as bcl2 may increase the sensitivity of detection of memory B, T cells.

CONCLUSION

Immunological memory is an important characteristic of the immune response and it differs from neurological memory in various respects. Various studies have characterized memory B and T cells according to lineage, longevity, circulation pattern and phenotypic markers. The ability to know how to detect the presence of memory B or T cells is very useful for medical treatment and for research in biological and medical science. However, this will be successful only when the complete characteristics of memory B and T cells are known and when reliable and convenient methods to assay these characteristics are developed.

ACKNOWLEDGEMENTS

Dr Timothy W Flagel's reading and Mr Charnchai Champreecha's typing are appreciated.

REFERENCES

- MacDonald HR, Budd RC, Cerottini JC. Pgp-1 (Ly 24) as a marker of murine memory T lymphocytes. Curr Top Microbiol Immunol 1990; 159 97-106.
- Zinkernagel RM. Antiviral T-cell memory? Curr Top Microbiol Immunol 1990; 159: 65-74.
- Gray D, Leanderson T. Expansion, selection and maintenance of memory B-cell clones. Curr Top Microbiol Immunol 1990; 159: 1-14.
- Vitetta ES, Berton MT, Burger C, Kepron M, Lee OUT, Yin XM. Memory B and T cells. Ann Rev Immunol 1991; 9:193-217.
- Klinman NR, Linton PJ. The generation of B cell memory: A working hypothesis. Curr Top Microbiol Immunol 1990; 159: 19-33.
- Mackay CR. Immunological memory. Adv immunol 1993; 159: 19-65.

- Berek C, Ziegner M. The maturation of the immune response. Immnol Today 1993; 14:400-4.
- Kepler TB, Perelson AS, Cyclic re-entry of germinal center B cells and the efficiency of affinity maturation. Immunol Today 1993; 14:412-5.
- Dell CL, Lu YX, Claffin JL. Molecular analysis of clonal stability and longevity in B cell memory. J Immunol 1989; 143: 3364-70.
- Alexander GB, Michael SN, Cesar M. Discriminating intrinsic and antigen-selected mutational hotspots in immunoglobulin V genes. Immunol Today 1993; 14: 405-11.
- Beverley PCL. Human T-cell memory. Curr Top Microbiol Immunol 1990; 195: 111-19.
- Adelstein S, Pritchard-Briscoe H, Loblay RH, Basten A. Suppressor T cell memory. Curr Top Microbiol Immunol 1990: 159: 123-37.
- Mackay CR. T cell memory: the connection between function, phenotype and migration pathway. Immunol Today 1991; 12:189-92.
- Helbert MR, L'age-Stehr J, Mitchison NA. Antigen presentation, loss of immunological memory and AIDS, Immunol Today 1993; 14: 340-3.
- Birkeland ML, Johnson P, Trowbridge IS, Pure E. Changes in CD45 isoform expression accompany antigen-induced murine T-eell activation. Proc Natl Acad Sci USA 1989; 86:6734.
- Gray D. Regulation of immunological memory. Curr Opinion Immunol 1994; 6:425-30.
- Antonio AF, Benedita BR. Lymphocyte lifespans : homeostasis, selection and competition. Immunol Today 1993; 14: 25-9.
- Buckton KE, Court Brown WM, Smith PG. Lymphocyts survival in men treated with X-rays for ankylosing spondylitis. Nature 1967; 214: 470-3.
- Mackay CR, Marston WL, Dudler L. Naive and memory T cell show distinct pathways of lymphocyte recirculation. J Exp Med 1990; 171:801-17.
- 20. Ferguson TA, Mizutant H, Kupper TS.

Two integrin-binding peptides abrogate T cell-mediated immune responses *in vivo*. Proc Natl Acad Sci USA 1991; 88: 8072-6.

- Tamatani T, Kotani M, Tanaka T, Miyasaka M. Molecular mechanisms underlying lymphocyte recirculation. II. Differential regulation of LFA-1 in the interaction between lymphocytes and high endothelial cells. Eur J Immunol 1991; 21: 855-8.
- Westermann J, Persin S, Matyas J, van der Meide P, Pabst R. Migration of so-called naive and memory T lymphocytes from blood to lymph in the rat. The influence of IFN-gamma on the circulation pattern. J Immunol 1994; 152: 1755-50.
- Yednock TA, Rosen SD. Lymphocyte homing. Adv Immunol 1989; 44: 313-371.
- MacLennan ICM, Liu YJ, Oldfield S, Zhang J, Lane PJL. The evolution of B-cell clones. Curr Top Microbiol Immunol 1990; 159: 37- 64.
- Sprent J. T and B memory cells. Cell 1994, 76: 315-22.
- Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration : the multistep paradigm. Cell 1994; 76: 301-14.
- Hockenbery DM, Zutter M, Hickey W, Nahm M, Korsmeyer SJ. Bel 2 protein is topographically restricted in tissue characterized by apoptotic cell death. Proc Natl Acad Sci USA 1991; 88: 6961-5.
- Veis DJ, Sentman CL, Bach EA, Korsmeyer SJ. Expression of the Bcl-2 protein in murine and human thymocytes and in peripheral T lymphocytes. J Immunol 1993; 2546-54.
- Fernandez-Sarabia MJ, Biachoff JR. Bel-2 associates with the ras-related protein Rras p23. Nature 1993; 366:274-5.
- Nunez G, Hockenbery D, McDonnell TJ, et al. Bcl-2 maintains B cell memory. Nature 1991, 353:71-73.
- Nakayama KI, Nakayama K, Negishi I, et al. Disappearance of the lymphoid system in Bcl-2 homozygous mutant chimeric mice. Science 1993, 261:1584-1588.
- Akbar AN. Inhibition of alloresponsive naive and memory T cells by CD7 and CD25 antibodies and by cyclosporine. Transplantation. 1990; 50:823-9.

- Ellis TM, Simms PE, Slionik DJ, Jack MH, Fisher RI. CD30 is a signal-transducing molecule that defines a subset of human activated CD45 RO+T cell. J Immunol 1993; 151: 2380-9.
- Visser L, Lai R, Poppema S. Patterns of leucocyte common antigen expression in peripheral blood T cell populations. Cell Immunol 1993; 151:218-24.
- Akbar AN, Salmon M, Janossy G. The synergy between naive and memory T cell during activation. Immunol Today 1991;12:184-81.
- Schwihoffer T, Tanaka Y, Tidswell M, et al. Selective expression of integrin 47 on a subset of human CD4+ memory T cell with hallmark of gut trophism. J Immunol 1993, 151: 717-29.
- Dianzani U, Redoglia V, Bragardo M, et al. Co-stimulatory signal delivered by CD73 molecule to human CD45 RA hi CD45 RO lo (naive) CD8+ T lymphocytes. J Immunol 1993; 151: 3961-70.
- 38. Urbaniak SJ, McCann MC, White AG, et al. Handbook of experimental immuno-

logy volume 4. Applications of immunological methods in biomedical sciences. Edited by DM Weir, 4th ed, 1986, c 126.

- Festin R, Bjorkland A, Totterman TH. Single laser flow cytometric detection of lymphocyte binding three antibodies labelled with fluorescein, phycoerythrin and a novel tandem fluorochrome conjugate. J Immunol Methods 1990; 126: 69-78.
- Swain SL, Weinberg AD, English M. CD4+ T cell subsets, lymphokine secretion of memory cells and of effector cells that develop from precursors in vitro. J Immunol 1989; 144: 1788-99.
- Salmon M, Kitas GD, Bacon PA. Production of lymphokine mRNA by CD45R+ and CD45R- helper cells from human peripheral blood and by human CD4+ T cell clones. J Immunol 1989; 143: 907-801.
- Powers GD, Abbas AK, Miller RA. Frequencies of IL-2 secreting T cell in naive and antigen stimulated lymhocyte populations. J Immunol 1989; 140:3352.

- Shalaby MR, Palladin MA. Manual of clincal laboratory immunology. Edited by Rose NR, Friedman H, Fahey JL. 3rd ed, 1986; 300-3.
- Xu-Amano J, Aicher WK, Taguchi T. Selective induction of Th 2 cells in murine Peyer's patches by oral immunization. Int Immunol 1992; 4:433-45.
- Nordstrom I, Ferrua B. Reverse ELISPOT assay for clonal analysis of cytokine production. II. Enumeration of interleukin 1secreting cells by amplified avidin-biotin anti peroxidase assay. J Immunol Methods 1992; 150: 199-206.
- 46. Chen KS, Strober W. Cholera holotoxin and its B subunit enhance Peyer's patch B cell responses induced by orally administered influenza virus: disproportionate cholera toxin enhancement of IgA B cell response. Eur J Immunol 1990; 20: 433-6.
- 47. Witkowski JM, Li SP, Gorgas G, et al. Extrusion of the P glycoprotein substrate rhodamine-123 distinguishes CD4 memory T cell subsets that differ in IL-2 driven IL-4 production. J Immunol 1994; 153: 658-65.